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<u>L9</u>	L1 same screen\$	6	<u>L9</u>
<u>L8</u>	L5 same aldehyde same amino	2	<u>L8</u>
<u>L7</u>	L5 same (advantag\$ or useful\$)	6	<u>L7</u>
<u>L6</u>	L5 same plastic	0	<u>L6</u>
<u>L5</u>	L4 same (drug or medicine)	20	<u>L5</u>
<u>L4</u>	epoxide same coupl\$	1158	<u>L4</u>
<u>L3</u>	L1 same epoxide	0	<u>L3</u>
<u>L2</u>	L1 same aldehyde	0	<u>L2</u>
<u>L1</u>	(herb or arabidopsis) same plastic	84	<u>L1</u>

END OF SEARCH HISTORY

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L2: Entry 84 of 84

File: USPT

Sep 4, 1973

DOCUMENT-IDENTIFIER: US 3755895 A

TITLE: HERB SPOON

Brief Summary Text (2):

The spoon may be produced of metal, however also of plastic, resilient material, such as polypropylene. The advantage of the latter construction is that the present herb spoon can be produced entirely in one piece, the hinge being formed by a thin connecting strip between the two parts.

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L2: Entry 56 of 84

File: USPT

Feb 13, 1996

DOCUMENT-IDENTIFIER: US 5490351 A

TITLE: Low cost sod mat and method for propagation

Detailed Description Text (31):

FIG. 1 is a fragmentary cross section of a typical flower sod mat according to this invention. Reference Numeral 40 is a suitable surface on which to grow sod mats such as 4 mil black polyethylene film mulch or a porous polyethylene sheet such as VISPORE.RTM. manufactured by Tredigar in Richmond, Va. Other suitable surfaces include plywood, rubber sheeting or a concrete slab. In a field grown application the sod mat growing surface needs to prevent weeds from growing into and through the flower or plant sod mat from the soil below and also to encourage the flower roots to grow laterally and form a sod. In a flat grown application, the thin plastic film can serve to contain the roots and planting medium (Reference Numeral 44) in a flat with a very open bottom and also to make sod mat removal easier at harvest time. Reference Numeral 42 represents the polyolefin sod reinforcement. The polyolefin sod reinforcement is a pattern bonded, polyolefin nonwoven fabric. A preferable example is a polypropylene sod reinforcement consisting of a polypropylene spunbond fabric such as CELESTRA.RTM. marketed by Fiberweb North America Inc. in Greenville, S.C. or ACCORD.RTM. marketed by Kimberly-Clark in Neenah, Wis. Reference Numeral 44 is the planting medium. The planting medium is adjusted to the optimum depth for the herb, vegetable, groundcover, flower, or plant species of sod mat being grown. FAIRGROW.RTM. is a good growing medium manufactured by Delaware Solid Waste Authority in Wilmington, Del. Reference Numeral 46 represents the demolition woodchips manufactured by Corrado American in Wilmington, Del. which can be added to the planting medium (Reference Numeral 44). Other planting medium amendments are also effective such as PERLITE.RTM., straw, hay, vermiculite, and are well known to those skilled in the art. Many other planting mediums can also be used. Examples include but are not limited to potting soil, METRO-MIX.RTM. and REDI-EARTH.RTM. manufactured by W. R. Grace. Reference Numeral 48 represents the seeds, seedlings, plant plugs, rooted cuttings, or root divisions added to the planting medium (Reference Numeral 44). Reference Numeral 48 also represents other means of starting plants where appropriate, such as cuttings and viable plant material derived from various types of plant tissue culture and thus are meant to be included in the seeds, seedlings, plant plugs, rooted cuttings, and root divisions definition above. Seedlings are usually planted in a 10-20 cm grid pattern. Thus various means to start plants are often referred to as plant starting materials in this specification and are well known to those of ordinary skill in the art (Reference Numeral 48). Reference Numeral 50 represents an optional crop cover such as REEMAY.RTM.2006 manufactured by Reemay, Inc., in Old Hickory, Tenn. This can serve to warm the soil up quickly in the spring and/or for some protection from marauding animals such as rabbits or birds.

Detailed Description Text (69):

In FIG. 5, there is shown a block diagram illustrating the general process of growing herb, vegetable, flower, groundcover or plant sod mats in a container according to this invention. The six generalized steps are: (1) the step 70 of selecting an appropriate flat for growing the herb, vegetable, flower, groundcover or plant sod. Generally the flat is from 5-10 cm deep, about 25-40 cm wide and 50-70 cm. long. Size is selected based on the depth the flowers or plants need to grow efficiently and on overall weight. Weight is generally less than 30 kgs. If a bottom suitable for roots to grow on is not inherent in the flat design, a separate layer of plastic film (such as VISPORE.RTM. or one of numerous black plastic film mulches)

is installed on the bottom. The plastic film then becomes Reference Numeral 40 and serves to contain the sod mat roots which penetrate the spunbond fabric installed in the next step. The plastic film also makes removing the sod mat at harvest time easier in many cases. If used, porous plastic films are generally preferred for flats. (2) the step 72 consists of installing the polypropylene sod reinforcement (Reference Numeral 42) on this plastic film or bottom of the flat (Reference Numeral 40). (3) step 74 consists of selecting and installing the planting medium (Reference Numeral 44) to the depth required of the herb, vegetable, groundcover, flower or plant species. (4) step 76 consists of adding the seeds, seedlings, plant plugs, rooted cuttings, or root divisions (Reference Numeral 48 ) of the desired species to the planting medium (Reference Numeral 44). (5) step 78 consists of normal feeding and care of the seeds, seedlings, rooted cuttings, root divisions or plant plugs (Reference Numeral 48) for maximum growth. Examples include maintaining proper moisture, shade, fertilizer, and soil amendments. MIRACLE-GROW.RTM. manufactured by Stern's MIRACLE-GROW.RTM. Products, Inc. in Port Washington, N.Y. is a good fertilizer when applied according to directions. One needs to take care not to over or underwater the medium. A flat which drains and uses porous polyethylene is often advantageous. (6) step 80 consists of harvesting the sod by removing the sod mat from the flat and selling as an herb, vegetable, groundcover, flower or plant sod mat. If a plastic film was used in the flat, it is also removed from the sod mat before final planting.

Detailed Description Text (101):

Accordingly, the reader will see that this invention can be used to economically and flexibly produce custom, high quality plant sod mats for the grower, landscaper and final customer. Many unique advantages are made available to these customers with this invention, such as excellent root penetration and entanglement, low equipment costs, economical small volume production costs, and high quality sod mats of many different herbs, vegetables, flowers and groundcovers. Furthermore multiple soil or soilless mediums are practical because of the unique flexibility of the polyolefin or polypropylene sod reinforcements discovered. Sod mats with viable seedlings, root divisions, rooted cuttings or plant plugs give instant beautification and make installation easier for the landscaper or homeowner. In addition, unique advantages are offered to both environment and customers by reducing packaging waste, reducing synthetic plastic reinforcement consumption, reducing the synthetic reinforcement planted with the sod along with the added advantage of having a reinforcement which will decompose. Useful decomposition can occur from ultraviolet light, biodegradation, and oxidation or a combination of these. The strength of the sod mat is tailored to the grower's and landscaper's requirements. Very low tear strength polypropylene spunbond fabrics have been shown to make excellent commercial size flower sod mats which are also easily divisible by hand with minimum root damage. The grower receives still more advantages by reducing his need for inventory, storage, disposal and handling costs associated with a complex array of planting pots and flats by using some simple rolls of polyolefin or polypropylene sod reinforcements which serve multiple plant species.

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L9: Entry 2 of 6

File: USPT

May 14, 2002

DOCUMENT-IDENTIFIER: US 6388089 B1

TITLE: Brassinosteroid biosynthesis inhibitors

Detailed Description Text (81):

The Arabidopsis germination assay was performed as follows. Seeds of wild-type and det were treated at a low temperature (2.degree. C.) for 2 days, and the surfaces were sterilized with 1% NaOCl solution for 2 minutes and washed seven times with sterilized distilled water. The seeds were sown on 1% agar solid medium containing 0.5.times.Murashige and Skoog (1962) salt and 1.5% (w/v) sucrose in a plastic plate in the presence or absence of the test compound. The wild-type plant and det2 plant were grown in a growth chamber for 16 hours under a light condition (240 mE/m2s) and for 8 hours under a dark condition (25.degree. C.). For the screening experiment, the plate was sealed with Parafilm (American National Can Co., Ltd., Chicago, Ill., USA). For the restoration experiment that required a longer experimental period, the seeds were sown on 1% agar solid medium containing 0.5.times.Murashige and Skoog (1962) salt and 1.5% (w/v) sucrose in Agripot (Kirin Brewery Co., Ltd.). The plants were grown in a growth chamber for 16 hours under a light condition (240 mE/m2s) and for 8 hours under a dark condition (28.degree. C.).

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L7: Entry 2 of 6

File: USPT

Mar 27, 2001

DOCUMENT-IDENTIFIER: US 6207369 B1

TITLE: Multi-array, multi-specific electrochemiluminescence testing

Detailed Description Text (44):

The porous material may be able to support a current due to the flow of ionic species. In a further refinement, the porous material is a porous water-swollen gel, for example polyacrylamide or agar. A variety of other gel compositions are available (for example see Soane, D. S. *Polymer Applications for Biotechnology*; Soane, D. S., Ed.; Simon & Schuster: Englewood Cliffs, N.J., 1992 or *Hydrogels in Medicine and Pharmacy*, Vol. I-III; Peppas, N. A. Ed.; CRC Press: Boca Raton, Fla., 1987). Binding domains can be attached to matrices by covalent and non-covalent linkages. (Many reviews and books on this subject have been written; some examples are Tampion J. and Tampion M. D. *Immobilized Cells: Principles and Applications* Cambridge University Press: NY, 1987; *Solid Phase Biochemistry: Analytical and Synthetic Aspects* Scouten, W. H. Ed., John Wiley and Sons: NY, 1983; *Methods in Enzymology, Immobilized Enzymes and Cells*, Pt. B Mosbach, K. Ed., Elsevier Applied Science: London, 1988; *Methods in Enzymology, Immobilized Enzymes and Cells*, Pt. C Mosbach, K. Ed., Elsevier Applied Science: London, 1987; *Methods in Enzymology, Immobilized Enzymes and Cells*, Pt. C Mosbach, K. Ed., Elsevier Applied Science: London, 1987; see also *Hydrogels in Medicine and Pharmacy*, supra). For example, a protein can be attached to a cross linked copolymer of polyacrylamide and N-acryloylsuccinimide by treatment with a solution of the protein. The binding domains may also be integrated into a porous matrix in a step prior to polymerization or gelation. In one embodiment, binding domains may be attached to uncrosslinked polymers by using a variety of coupling chemistries. The polymers may then be crosslinked (for example using chemistries which include amide bonds, disulfides, nucleophilic attack on epoxides, etc.) (see for example: Pollack et al., 1980, J. Am. Chem. Soc. 102(20):6324-36). Binding domains may be attached to monomeric species which are then incorporated into a polymer chain during polymerization (see Adalsteinsson, O., 1979, J. Mol. Catal. 6(3): 199-225). In yet another embodiment, binding domains may be incorporated into gels by trapping of the binding domains in pores during polymerization/gelation or by permeation of the binding domains into the porous matrix and/or film. Additionally, binding domains may be adsorbed onto the surface of porous matrices (e.g., polymer gels and films) by nonspecific adsorption caused for example by hydrophobic and/or ionic interactions. Biotin may be advantageously used as a linking or binding agent. Avidin, streptavidin or other biotin binding agents may be incorporated into binding domains.

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L9: Entry 4 of 6

File: USPT

Sep 30, 1980

DOCUMENT-IDENTIFIER: US 4224743 A

TITLE: Food dehydrating machine

Brief Summary Text (12):

An additional feature of the present invention is the use of trays including supporting sheets for the foods. The supporting sheets may be screens of various mesh sizes depending upon the food to be dried. If coarse food pieces such as carrots are being dried, coarse screen sheet may be used, for example, one-half inch grid. If fine food pieces such as herbs are being dried, fine screen may be used. Alternatively, if desired, the food supporting trays can be made of a solid sheet. This is particularly suitable if the food being treated is originally in a paste or slurry state. The supporting sheets may be removably mounted in the trays. The supporting sheets may be of any food safe material such as stainless steel or plastic. The sheets desirably provide a minimum of sticking. The supporting sheets are removably mounted in the tray frames so that they may be removed and flexed to pop off any food chips which are stuck to the sheet. The removability feature also provides for interchanging of sheets having various opening sizes. The sheet can be removed from the frame and moved into funnel-like shape while the contents are poured into a container. It is desirable to use support sheets having the largest openings which still serving to support the food pieces.

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L8: Entry 1 of 2

File: USPT

May 8, 2001

DOCUMENT-IDENTIFIER: US 6229055 B1

TITLE: Synthesis of fluorinated xanthene derivatives

Detailed Description Text (400):

Specific embodiments of the dyes of the present invention that possess particular utility for the preparation of dye-conjugates are amino, hydroxy and carboxy derivatives of the fluorinated fluorophores, as these derivatives are readily converted to a wide variety of other reactive derivatives. These reactive derivatives in turn can be conjugated to amino acids, peptides, proteins, nucleotides, oligonucleotides, nucleic acids, carbohydrates, polysaccharides, lipids, drugs, toxins, ligands and other molecules of interest using standard chemistry that is typically accomplished at or near room temperature. For instance, amine derivatives are converted to isocyanates, isothiocyanates, ureas, thioureas, urethanes, semicarbazides, dichlorotriazinyl amines, amides, haloacetamides, maleimides, acrylamides, azides, hydrazines or alkylated amines; carboxylic acids are converted to esters or activated esters capable of forming amides or esters, amides, hydrazides, acid halides, acyl azides, acyl nitrites or reduced to aldehydes or alcohols; hydroxy groups are esterified, alkylated to ethers including glycosides, or converted to alkylating agents such as halogens or sulfonate esters. Other synthetic strategies for the preparation of useful reactive derivatives include the formation of aldehydes, glyoxals, thiols, epoxides, aziridines, sulfonyl halides, imido esters, borates or phosphoramidites. Alkenes for use in polymerization can be incorporated by various means, including reacting with functionalized styrenes or allylamines or by reaction of an amine on the dye with an acrylic acid derivative. All of these reactions are accomplished using methods well known in the art. The reactions that are typically used to couple the reactive fluorophores of the invention with molecules of interest include, but are not limited to, those listed in Table 2.



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L2: Entry 10 of 84

File: USPT

May 28, 2002

DOCUMENT-IDENTIFIER: US 6395963 B1

TITLE: Nematode-inducible regulatory DNA sequences

Detailed Description Text (42):

Arabidopsis seeds were surface sterilized and sown in petri dishes (.phi.:9 cm) on B5 medium containing 20 g/l glucose and 20 mg/l kanamycin. After 3 days at 4.degree. C. the plates were incubated for 2 weeks in a growth chamber at 22.degree. C. with 16-hr light/8 hr-dark cycle. Kanamycin-resistant plants were then transferred to soil-filled translucent plastic tubes (30.times.15.times.120 mm, Kelder plastibox b.v., The Netherlands). The tubes were placed tilted at an angle of 60 degrees to the vertical axis causing the roots to grow on the lower side of the tubes. This allows to monitor the infection process by eye and facilitates removal of the root system from the soil for GUS analysis. Infection was done after two more weeks by injecting a suspension containing 500 second stage larvae of *Heterodera schachtii* (in 3 ml H.sub.2 O) per root system or 300 second stage larvae of *Meloidogyne incognita* per root system into the soil.

Detailed Description Text (56):

The binary vector pMOG553 was mobilized by triparental mating to *Agrobacterium tumefaciens* strain MOG101 which is described in detail in WO 93/10251. The resulting strain was used for Arabidopsis root transformation. More than 1100 transgenic Arabidopsis plant lines were obtained in this way. Transgenic plants were grown to maturity, allowed to self-fertilize and the resulting seeds (S1) were harvested and vernalized. Subsequently S1 seeds were germinated on nutrient solution (Goddijn et al. 1993 Plant J 4, 863-873) solidified with 0.6% agar, 10 mg/l hygromycin and stored at 4.degree. C. for a 4 day imbibition period. At day 5 the plates were transferred to room temperature and moderate light (1000 lux, 16 h L/8 h D) for germination. Fourteen days old seedlings were transferred to potting soil in tilted translucent plastic tubes (30.times.15.times.120 mm) for further growth at 5000 lux (20.degree. C.). Growing the plants in this way causes most of the root system to grow on the lower side of the tubes in the interphase between soil and tube. After two weeks the roots were infected with nematodes as described in the Experimental part. At several time points after inoculation (ranging from 2-14 days), the root systems were analyzed for GUS activity as described in the Experimental part. Line pMOG553#25 was identified as a line which showed rather strong GUS expression inside syncytia and giant cells induced by *Heterodera schachtii* and *Meloidogyne incognita*, respectively. In uninfected control plants (as well as in the infected plants) of this line very weak GUS expression was detected in the vascular cylinder at the base and at the tip of young lateral roots and in various green parts of the plant.

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L2: Entry 31 of 84

File: USPT

Apr 10, 2001

DOCUMENT-IDENTIFIER: US 6214350 B1

TITLE: Process for preparing an anti-viral medicinal product from plant extracts

Detailed Description Text (234):

A 5.0 g sample of each single-herb herbal medicine No.4(2), No.4(3), No.4(4), No.4(5), No.5(1), No.5(4) and No.5(8) was extracted with about 40 mL water twice in a 50-mL plastic centrifuge tube. Each extract was separated from the insoluble material by centrifuge at 2,000 rpm for 40 to 120 minutes. The first and second extracts of each sample were combined and then filtered through a 0.22- $\mu$ m filter. A 2.00 mL aliquot of each extract was nitrogen dried and weighed. The remaining extract of each sample was acidified with 10 mL 1% HCl in water. Precipitates formed in all acidified extracts, except those of No.4(3) and No.5(4). No precipitate formed in the acidified extract of No.4(3), even after prolonged (9 hrs) storage in a refrigerator and addition of 10 mL more 1% HCl in water. The acidified extract of No.5(4) showed only cloudiness and formed a trace precipitate after centrifuge at 2,000 rpm for 20 minutes.

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L2: Entry 45 of 84

File: USPT

Jan 12, 1999

DOCUMENT-IDENTIFIER: US 5858446 A

TITLE: Processed fresh herbs and method of making

Detailed Description Text (21):

After mixing, the herb mix is packaged in sealed glass jars or other low oxygen transmission packages such as flexible laminates, metal or plastic tubes etc. In smaller containers for consumer use it is preferred that the type of package be such that the contents can be squeezed out and re-sealed (e.g. a metal tube) so that air is kept to a minimum within the package. The packaged product is held at low temperature, preferably between 8.degree. C. and -20.degree. C. and more preferably at -20.degree. C.

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L2: Entry 54 of 84

File: USPT

Jun 4, 1996

DOCUMENT-IDENTIFIER: US 5523102 A

TITLE: Method of improving the firmness of fish tissue

Detailed Description Text (21):

Fresh and frozen/thawed whiting fillets were pretreated and heated as described in Example 1. The tissue was firm. The whiting fillets were coated with Splash French Herbs Cooking Sauce.TM., stored in plastic bags, and frozen for three months. After being thawed and reheated, the fillets had nice appearance and a firm, palatable texture.